0040-4020(94)01032-3

Enantiopure Hydroxylactones from L-Ascorbic and D-Isoascorbic Acids. Part I.¹ Synthesis of (-)-Muricatacin

Michèle Sanière, Isabelle Charvet, Yves Le Merrer, Jean-Claude Depezay

Université René Descartes, Laboratoire de Chimie et Biochimie Pharmacologiques et Toxicologiques, associé au CNRS, 45, rue des Saints Pères, 75270 Paris Cedex 06, France.

Key-words. (-)-Muricatacin, D-isoascorbic acid, epoxylactone, 5-hydroxy- γ -butyrolactone, Mitsunobu reaction, bis-epoxide.

Abstract. From *D*-isoascorbic acid, *via* a formal bis-epoxide equivalent with a C-2 axis of symmetry, two possible syntheses of the (-)-Muricatacin are described.

Compounds with chiral hydroxylactone occupy important positions both as target bioactive molecules and useful synthetic equivalents in total syntheses. For example, natural 5-hydroxy-γ-lactones were identified as flavour constituents in wine,² sherry,³ and tobacco smoke⁴ and as a microbial metabolite in cultures of *Erwinia quernica*.⁵ The isolation of 5-hydroxy-γ-decalactone (L-Factor) from cultures of *Streptomyces griseus*⁶ which reveals autoregulatory properties and of 5-hydroxy-γ-heptadecalactone (Muricatacin) from seeds of *Annona muricata*, ⁷ an acetogenic derivative which shows some cytotoxicity on human tumour cell lines, has stimulated great interest and has been at the origin of synthetic strategies towards these products.^{8,9}

Our general approach to enantiomerically pure hydroxy- γ -butyro and δ -valerolactones starts either from L-ascorbic or D-isoascorbic acids (Scheme 1). As previously described, 10 these commercial acids are converted in 40 % overall yield into each of the four possible stereoisomers of epoxybutanediol acetonide 2. These epoxybutanediol acetonides are formal equivalents of bis-epoxide containing a free epoxide function, the other one being masked into the glycol; successive regiospecific nucleophilic openings of both epoxide rings allow the introduction of the alkyl chain, on one hand and the formation of the lactone ring, on the other hand.

1654 M. SANIÈRE et al.

Following this strategy, we recently proposed a route to (-)-Muricatacin, [(4R,5R)-5-hydroxy-4-heptadecanolide 1] and (-)-(5R,6S)-6-acetoxy-5-hexadecanolide, 1,9i and the present report provides the details of that effort and describes a new route to (-)-Muricatacin.

In the specific case of the (-)-Muricatacin (Scheme 2) which has a *threo* relative configuration, this strategy requires a formal bis-epoxide with a C-2 axis of symmetry. We take advantage of this two fold axis to study two approaches (path a and path b) from the same epoxybutanediol acetonide, these two paths differ only by the order of introduction of nucleophiles.

Scheme 2

C-2 axis of symmetry

D-Isoascorbic acid

path a

$$:Nu^{1} = : C_{11}H_{23}$$

$$:Nu^{2} = : CH(CO_{2}Et)_{2}$$

$$:Nu^{2}$$

$$:Nu^{2}$$

$$:Nu^{2}$$

$$:Nu^{3}$$

$$:Nu^{4} = : C_{11}H_{23}$$

$$:Nu^{2} = : CH(CO_{2}Et)_{2}$$

$$:Nu^{4}$$

$$:Nu^{2}$$

$$:Nu^{4}$$

$$:Nu^{2}$$

$$:Nu^{4}$$

$$:Nu^{5}$$

Path a: (Scheme 3)

The nucleophilic opening of (2R,3R)-3,4-epoxy-1,2-O-methylethylidenebutane-1,2-diol 2, prepared from D-isoascorbic acid, 10 with undecylmagnesium bromide in the presence of Li₂CuCl₄ led to the alcohol 3 (80%) which was protected as a 4-methoxybenzylether 4 (NaH, DMF, imidazole, 4-methoxybenzylchloride, 93%). Acidic hydrolysis (AcOH-H₂O) of 4 gave the diol 5 which was transformed into epoxide 6 either by action of NaH in DMF/THF followed by addition of tosylimidazole 11 (42%) or in a higher yield (83%) by Mitsunobu reaction 12 (PPh₃, DIAD, 125 °C in vacuo).

Next step was the introduction of acetate functionality at the other epoxy site of the bis-epoxide equivalent. So, 6 was treated with ethylmalonate in presence of sodium ethoxide to afford a mixture of α -carbethoxy- γ -butyrolactones epimers 7 in 55 % yield. Smooth decarbethoxylation of this crude mixture by magnesium chloride hexahydrate in dimethylacetamide¹³ followed by deprotection of the alcohol by dicyanodichloroquinone oxydation¹⁴ of its paramethoxybenzyl protecting group led to the expected (-)-Muricatacin 1 in 71 % yield.

a) $C_{11}H_{23}MgBr$, $L_{12}CuCl_4$, THF, $-35^{\circ}C$ from 2 (80 %); and $-78^{\circ}C$ from 16 (58%). b) NaH, DMF, imidazole, 0°C then MPMCl, NBu₄I, 20°C, 93 %. c) AcOH/H₂O 4/1, 20°C overnight: quantitative yield from 4 and 3 respectively. d) PPh₃, DIAD, 125°C *in vacuo*, 83 % from 5, 70 % for $3 \rightarrow 9 \rightarrow 10$, 67 % from 13. e) CH₂(COOEt)₂, EtOH-EtONa, 60°C, 6 hrs, 55 %, 80 % from 6, 2 respectively. f) MgCl₂.6H₂O, CH₃CON(CH₃)₂, reflux 4 hrs, 90 % and 20 % overall yield from 12 and 10 respectively. g) DDQ, CH₂Cl₂/H₂O, 71 %. h) TsCl 1 eq, NEt₃ 1.5 eq, CH₂Cl₂, $-20^{\circ} \rightarrow 20^{\circ}C$, 26 %. i) TIPSCl 1.3 eq, pyridine, 0° $\rightarrow 20^{\circ}C$. 41 %. j) NaH, THF/DMSO 0° $\rightarrow 20^{\circ}C$, 2 hrs, 15 \rightarrow 16, 63 %.

DIAD: diisopropyl azadicarboxylate; MPMCl: 4-methoxybenzylchloride. TIPSCl: triisopropyl benzenesulfonyl chloride. DDQ: dicyanodichloroquinone.

1656 M. SANIÈRE et al.

An alternative way to (-)-Muricatacin from the acetonide alcohol 3 without protection of the hydroxyl group was tested. The 1,2-epoxy-3-alkanol 10 was obtained from the acyclic triol 9 in 70 % yield according to Mitsunobu conditions. ¹⁵ However its nucleophilic opening following by decarbethoxylation afforded the (-)-Muricatacin in 20 % from the epoxide 10 compared to 40 % in the protected way.

Path b: (Scheme 3)

Now, nucleophilic opening of the epoxide 2 with diethylmalonate in the presence of sodium ethoxide afforded a mixture of α -carbethoxy- γ -butyrolactone diastereomers 12a and 12b (70/30) in 80 % yield. Magnesium chloride hexahydrate in refluxing dimethylacetamide induced decarbethoxylation simultaneously with hydrolysis of the acetonide to afford 13 in 90 % yield. The selective activation of primary alcohol with tosylchloride (Et₃N, CH₂Cl₂) or triisopropylbenzenesulfonyl chloride in pyridine, was carried out with low yields; 14 and 15 were obtained in 26 % and 41 % yield respectively and 15 was transformed (NaH, Me₂SO) in epoxylactone 16 in 63 % yield. Nevertheless, Mitsunobu reaction 12 (PPh₃, DIAD, 125°C *in vacuo*) achieved on the diol lactone 13 led directly to the epoxylactone 16 in 65 % yield.

Finally, the nucleophilic opening of the epoxylactone 16 by the undecylmagnesium bromide in presence of Li₂CuCl₄ led to (-)-Muricatacin 1 in 38 % yield together with 34 % of the starting material which could be easily recovered by flash column chromatography (58 % yield of 1 based on 66 % conversion of 16).

The butyrolactones are known to exist in the envelope conformation and in solution an equilibrium of two conformers must be considered (scheme 4). 16 NMR study of 1 in CDCl₃ (see experimental section) shows that the (-)-Muricatacin takes up a predominant conformation with the 4-hydroxyalkyl chain in equatorial position. Thus, the expected values for $^3J_{3,4}$ (ax,ax) and $^3J_{3,4}$ (eq,eq) proton coupling should be 10 Hz and 2 Hz respectively. 16b For the lactone 1 the experimental value ($^3J_{3,4}$ experimental error 0.5 Hz) may be interpreted as a slight contribution of 1b which is evaluated as 25% by the equation: $2(1-\alpha) + 10\alpha = 8$ (α is the molar concentration of configuration 1a).

Scheme 4

$$H_{3\beta}$$
 $H_{3\alpha}$
 $H_{2\beta}$
 H_{4}
 $H_{2\alpha}$
 $H_{2\alpha}$
 $H_{2\alpha}$
 $H_{2\alpha}$
 $H_{3\alpha}$
 $H_{3\alpha}$
 $H_{3\alpha}$
 $H_{3\alpha}$
 $H_{3\alpha}$
 $H_{3\alpha}$

In summary, we have described two routes to (-)-Muricatacin from D-isoascorbic acid via an epoxy butanediol acetonide, a formal bis-epoxide equivalent with a C-2 axis of symmetry. One of them, via an epoxy-putyrolactone ($path\ b$), involves only four steps without protection-deprotection sequences, and allows the introduction of the alkyl chain in an ultimate step of the synthesis.

A key feature of this method is that the enantiomer of the epoxybutanediol acetonide can also be obtained from L-ascorbic acid and therefore allows access to (+)-Muricatacin.

Furthermore, generalisation of this versatile procedure to other nucleophiles could also lead to a variety of enantiomerically pure 5-hydroxy-y-butyrolactones such as:`

- (4S, 5S)-5-hydroxy-4-decanolide (:Nu=C₄H₉), L-Factor.⁶
- (4S,5S)-5-hydroxy-4-pentadecanolide (:Nu=C₉H₁₉), an useful building block for the synthesis of disparlure.¹⁷
- (4R,5R)-5-hydroxy-7-phenyl-4-hexanolide (:Nu=C₆H₅), a microbial metabolite in culture of *Erwinia quercina*.⁵

EXPERIMENTAL SECTION

Prior to use, tetrahydrofuran (THF) and diethyl ether (Et₂O) were distilled from sodium-benzophenone and dichloromethane (CH₂Cl₂) from P₂O₅. CH₂Cl₂ and ethyl acetate (AcOEt) were filtered on K₂CO₃ prior to use. 1 H NMR (250 MHz) and 13 C NMR spectra were recorded in CDCl₃ (unless indicated) on a Bruker AM 250. Chemical shifts are reported in δ (ppm) and coupling constants are given in Hertz. High Resolution Mass Spectra were recorded in Service de Spectrométrie de Masse, Université Pierre et Marie Curie. Specific rotations were measured on a Perkin Elmer 241C polarimeter with sodium (589 nm) or mercury (365 nm) lamps. All reactions were carried out under argon atmosphere, and were monitored by thin-layer chromatography with Merck 60F-254 precoated silica (0.2 mm) on glass. Chromatography was performed with Merck Kieselgel 60 (200-500 μ m) or 60H (5-40 μ m). Spectroscopic (1 H and 13 C NMR, MS) and/or analytical data were obtained using chromatographically homogeneous samples.

(2R, 3R)-1,2-O-Methylethylidene-pentadecane-1,2,3-triol (3)

To a stirred solution of Li₂CuCl₄ (3.19 mmol, 0,1M in THF prepared from 1 mol. CuCl₂ and 2 mol. LiCl₃ at -35°C, undecylmagnesium bromide (31.9 mmol, 1M in THF prepared from magnesium turnings and 1-bromo undecane in refluxing THF for 30 min)¹⁸ was added dropwise. After stirring for 30 min. at -35°C, epoxide 2 (1.044g, 7.25 mmol) in THF (42 mL) was added and the mixture was stirred 30 min. at -35°C. Hydrolysis with a saturated aqueous solution of ammonium acetate was followed by extraction with ether (4x75 mL). The combined ether layers were washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. Flash chromatography of the residue (cyclohexane/AcOEt 90:10 Rf 0.16) afforded 1.66g (80 %) of 3:

[α]_D +12.4 (c 1.11, CH₂Cl₂); ¹H NMR δ : 3.97 (2H, m, H-1), 3.70 (1H, m, H-2), 3.46 (1H, m, H-3), 1.41, 1.35 (6H, 2s, CMe₂), 1.1-1.45 (22H, m, (CH₂)₁₁), 0.85 (3H, t, CH₃, J=6.5Hz); ¹³C NMR δ : 109.3 (CMe₂), 79.2 (C-2), 72.3 (C-3), 66.2 (C-1), 33.7 (C-4), 31.9, 29.6, 29.3, 25.5, 22.7 (C-5-14), 26.7, 25.3 (CMe₂), 14.1 (C-15). Anal. Calcd. for C₁₈H₃₆O₃ : C, 71.95 ; H, 12.08. Found : C, 72.06 ; H, 11.99,

(2R, 3R)-1,2-O-Methylethylidene-3-O-para-methoxybenzyl-pentadecane-1,2,3 triol (4)

To a stirred suspension of NaH (148 mg, 6.16 mmol) in DMF (5.2 mL) at 0°C was added the alcohol 3 (463 mg, 1.54 mmol) in DMF (3.7 mL) and an imidazole crystal. After stirring 2 hrs at room temperature paramethoxybenzylchloride (733 μ L, 5.4 mmol) was added dropwise. The mixture was stirred overnight then poured into water and extracted with ether (4x15 mL). The combined ether layers were washed with brine,

dried (MgSO₄), filtered and concentrated *in vacuo*. Flash chromatography of the residue (cyclohexane/AcOEt 90:10, Et₃N : 3 % Rf 0.29) afforded 603 mg (93 %) of 4 :

[α]_D +29 (c 2.73, CH₂Cl₂); ¹H NMR δ : 7.28, 6.87 (4H, 2d, Ar-H, J=8Hz), 4.64, 4.54 (2H, 2d, CH₂-Ar, ²J=11Hz), 4.19 (1H, ddd, H-2, J_{1,2}=7, J_{1',2}= J_{2,3}=6.5Hz), 3.97 (1H, dd, H-1, J_{1,1}=8.2Hz, J_{1,2}=7Hz), 3.8 (3H, s, Ar-OCH₃), 3.66 (1H, dd, H-1', J_{1,1}=8.2Hz, J_{1',2}=6.5Hz), 3.39 (1H, m, H-3), 1.44, 1.37 (6H, 2s, CMe₂), 1.55-1.05 (22H, m, (CH₂)₁₁), 0.88 (3H, t, CH₃, J=6.5Hz). ¹³C NMR δ : 159.2, 131.0, 129.5, 113.7 (Ar), 109.2 (\underline{C} Me₂), 79.4 (C-2), 78.5 (C-3), 72.5 (\underline{O} CH₂), 66.0 (C-1), 55.2 (Ar-OCH₃), 31.9, 30.7, 29.7, 29.3, 25.6, 22.7 (C-4-14), 26.6, 25.5 (CMe₂).

(2R, 3R)-3-*O-para*-Methoxybenzyl-pentadecane-1,2,3-triol (5)

The acetonide **4** (603 mg, 1.44 mmol) in CH₃COOH-H₂O 4-1 (10.3 mL) was stirred overnight at room temperature. The mixture was concentrated *in vacuo* to give an oil which was purified by silicagel chromatography (CH₂Cl₂/MeOH 95:5, Et₃N 3% Rf 0.25) to give diol **5** (75 %): m.p.=45°C,

[α]_D -21 (c 1.01, CH₂Cl₂); ¹H NMR δ : 7.28, 6.87 (4H, 2d, Ar-H, J=8Hz), 4.60, 4.39 (2H, 2d, CH₂-Ar, ²J=11Hz), 3.8 (3H, s, Ar-OCH₃), 3.63 (3H, m, H-1-2), 3.46 (1H, m, H-3), 1.58 (2H, m, H-4), 1.4-1.2 (20H, m, (CH₂)₁₀), 0.88 (3H, t, CH₃, J=6.5Hz). ¹³C NMR δ : 159.4, 130.4, 129.4, 140.0 (Ar), 79.3 (C-3), 73.0 (C-2), 71.9 (OCH₂Ar), 64.1 (C-1), 55.2 (Ar-OCH₃), 31.9 (C-4), 30.3, 29.8, 29.6, 29.3, 25.2, 22.6 (C-5-14), 14.0 (C-15).Anal. Calcd. for C₂₃H₄₀O₄ : C, 72.59 ; H, 10.59. Found : C, 72.46 ; H, 10.58.

(2R, 3R)-1,2-Epoxy-3-*O*-para-Methoxybenzyl-3-pentadecanol (6)

At 0°C, diisopropyl azodicarboxylate (DIAD, 338 μ L, 1.71 mmol) was added dropwise to a stirred solution of diol 5 (496 mg, 1.3 mmol) and triphenylphosphine (443 mg, 1.69 mmol) in dry benzene (2 mL) (diol and triphenylphosphine were previously concentrated twice *in vacuo* from a toluene solution to avoid any trace of water). After stirring for 30 min at 0°C, the benzene was removed *in vacuo* and the residue was heated to 130°C (0.03 mm Hg) for 2 hrs. Flash chromatography of the residue (CH₂Cl₂, NEt₃ 3‰, Rf 0.25) afforded 395 mg (83 %) of 6 :

 $[α]_D$ +20 (c 1.02, CH₂Cl₂); 1 H NMR δ : 7.28, 6.85 (4H, 2d, Ar-H, J=8Hz), 4.75, 4.5 (2H, 2d, CH₂-Ar, 2 J=11Hz), 2.99 (2H, m, H-2, H-3), 2.75 (1H, m, H-1), 2.47 (1H, m, H-1'), 1.2-1.5 (22H, m, (CH₂)₁₁), 0.86 (3H, t, CH₃, J=6.5Hz). 13 C NMR δ : 159.0, 130.8, 129.3, 113.6 (Ar), 80.0 (C-3), 71.3 (OCH₂Ar), 55.1 (Ar-OCH₃, C-2), 43.1 (C-1), 32.3, 31.9, 29.6, 29.3, 25.5, 22.6 (C-5-14), 14.0 (C-15).

Anal. Calcd. for C23H38O3: C. 76.20: H. 10.56. Found: C. 76.11: H. 10.65.

(4R, 5R)-2-Carbethoxy-5-*O-para*-methoxybenzyl-4-heptadecanolide (7)

To a solution of EtONa [prepared from Na (24 mg, 1 mmol) in EtOH (750 μ L)] was added diethylmalonate (172 μ L, 1.1 mmol), followed by the epoxide 6 (205 mg, 0.56 mmol) in EtOH (650 μ L). After refluxing for 6 hrs, the mixture was poured into a saturated aqueous solution of NH₄Cl and extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. Flash chromatography of the residue (cyclohexane/AcOEt 80:20, NEt₃ 3% Rf 0.2) afforded 7 (55 %).

¹H NMR δ : 7.24, 6.85 (4H, 2d, ArH, J=8Hz), 4.64, 4.5 (2H, 2d, CH₂-Ar, ²J=11Hz), 4.5 (1H, m, H-4), 4.20 (2H, q, CH₂, J=7Hz), 3.80 (3H, s, Ar-OCH₃), 3.6 (1H, m, H-2), 3.48 (1H, m, H-5), 2.10 (2H, m, H-3), 1.46, 1.29 (27H, m, (CH₂)₁₂, CH₃), 0.84 (3H, t, CH₃, J=7Hz). ¹³C NMR δ : 171.4 (C-1), 167.5

(COOEt), 159.2, 130.3, 129.6, 129.4, 113.8 (Ar), 81.5 (C-4), 80.2, 79.7 (C-5 dia), 72.8, 71.8 (OCH₂Ar dia), 66.1 (OCH₂CH₃), 55.2 (Ar-OCH₃), 46.8 (C-2), 31.9, 29.9, 29.6, 29.3, 28.6, 25.4, 25.1, 22.7 (C-6-16), 14.1 (CH₃).

(4R, 5R)-5-O-para-Methoxybenzyl-4-heptadecanolide (8)

To a solution of 7 (71 mg, 0.15 mmol) in N,N-dimethylacetamide (590 µL) was added MgCl₂.6H₂O (152 mg, 0.75 mmol). The mixture was refluxed for 3 hrs with stirring. After cooling, the mixture was extracted with ether. The combined organic layers were washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. The crude product was used in the next step without further purification.

 1 H NMR (90 MHz) δ : 7.25, 6.90 (4H, 2d, ArH, J=8Hz), 4.5-4.3 (3H, m, OCH₂Ar, H-4), 3.8 (3H, s, Ar-OCH₃), 3.4 (1H, m, H-5), 2.7-2.3 (2H, m, H-2), 2.3-1.9 (2H, m, H-3), 1.7-1.0 (20H, m, (CH₂)₁₀), 0.9 (3H, t, CH₃, J=6.5Hz).

(2R, 3R)-Pentadecane-1,2,3-triol (9)

Acid hydrolysis of acetonide 3 by CH₃COOH-H₂O 4-1 was carried out under identical conditions as for 4 described above. Rf 0.09 (AcOEt/cyclohexane 7:3). The crude product was used in the next step without further purification.

¹H NMR (90 MHz) δ : 3.4-3.8 (4H, m, H-1-3), 1.32 (22H, m, (CH₂)₁₁), 0.90 (3H, t, CH₃, J=6.5Hz).

(2R, 3R)-1,2-Epoxy-3-pentadecaneol (10)

Mitsunobu reaction on 9 was carried out under identical conditions as for 5 described beforehand. Flash chromatography (cyclohexane/AcOEt 7:3, Rf 0.30) afforded 1 0 (70 % yield from 3).

[α]_D +2 (c 1.32, CH₂Cl₂), [α]_{Hg 365} +9 (c 1.32, CH₂Cl₂). ¹H NMR δ : 3.41 (1H, m, H-3), 2.96 (1H, m, H-2), 2.80 (1H, dd, H-1, 3 J_{1,2}= 2 J_{1,1}=4.5Hz), 2.69 (1H, dd, H-1', 2 J_{1,1}=4.5Hz, 3 J_{1',2}=3Hz), 1.56 (2H, m, H-4-4'), 1.24 (20H, m, (CH₂)₁₀), 0.83 (3H, t, CH₃, J=6.5Hz). ¹³C NMR δ : 71.3 (C-3), 55.0 (C-2), 44.8 (C-1), 34.0, 31.5, 29.2, 28.9, 24.9, 22.3 (C-4-14), 13.7 (CH₃).

(4R, 5R)-2-Carbethoxy-5-hydroxy-4-heptadecanolide (11)

Nucleophilic opening of the epoxide function by diethylmalonate on 10 was carried out under identical conditions as for 6 described beforehand. Rf 0.27 (cyclohexane/AcOEt 1/1). The crude product was used in the next step without further purification.

 1 H NMR (90 MHz) δ : 4.4 (1H, m, H-4), 4.2 (2H, q, OEt), 3.3-3.8 (2H, m, H-2, H-5), 2.2-2.8 (2H, m, H-3), 1.2-1.8 (22H, m, (CH₂)₁₁), 0.9 (6H, t, CH₃, J=6.5Hz).

(4R, 5R)-2-Carbethoxy-5,6-dihydroxy-5,6-O-methylethylidene-4-hexanolide (12)

Nucleophilic opening of the epoxide function by diethylmalonate on 2 was carried out under identical conditions as for 6 described beforehand. From the epoxide 2 (720 mg, 5 mmol) a mixture of 12a and 12b diastereomers (80 % yield, 70/30) was obtained after flash chromatography (CH₂Cl₂/Et₂O 75/25, Rf 0.6).

IR (film) 1780, 1735 cm⁻¹. ¹H NMR δ: 4.55 (1H, ddd, H-4 maj, J=2, 4.1, 9.9Hz), 4.44 (1H, ddd, H-4 min, J=4.2, 6.6, 8.3Hz), 4.25 (2H, q, O<u>CH</u>₂CH₃ min, J=6.6Hz), 4.26 (2H, q, O<u>CH</u>₂CH₃ maj, J=6.6Hz), 4.2-4.1 (2H, m, H-5 maj, H-5 min), 3.83-4.0 (4H, m, H-6, H-6' maj and min), 3.73 (1H, dd, H-2 maj, J=8.3, 9.9Hz), 3.09 (1H, t, H-2 min, J=9.9Hz), 2.79 (1H, ddd, H-3), 2.35-2.60 (3H, m, H-3 maj, H-

H-3, H-3' min), 1.2-1.4 (9H, 4m, CH₃). 13 C NMR δ : 172.0 (C-1), 168.0 (COOEt), 111.0 (CMe₂), 78.0, 77.4 (C-4 dia),76.3 (C-5), 65.2, 65.0 (C-6 dia), 62.1 (OCH₂CH₃), 46.2, 46.0 (C-2 dia), 28.9, 27.5 (C-3 dia), 26.1, 25.7, 25.4, 25.2 (CMe₂ dia), 14.0 (CH₂-CH₃). MS m/z (relative intensity) : 243 (M⁺-15 (100)), 213 (10), 155 (12), 109 (15), 137 (68), 101 (92). NH₃ chemical ionization 276 (M⁺+18), 259 (M⁺+1).

(4R, 5R)-5,6-Dihydroxy-4-hexanolide (13)

To a solution of 12 (776 mg, 3 mmol) in N,N-dimethylacetamide (12 mL) was added MgCl₂.6H₂O (3.05 g, 15 mmol). The mixture was refluxed for 4 hrs with stirring. After cooling, the mixture was poured in brine and washed with ether (5x50 mL). After acidification of aqueous layers until pH=3 and liophilisation, the residue was extracted with chloroform in a Soxhlet. Removal of solvent at reduced pression afforded 430 mg of 13 (90%) . TLC: CH₂Cl₂/MeOH 8/1, Rf: 0.22.

[α]_D -43.3 (c 0.9, CH₂Cl₂). IR (film) : 3380, 1775 cm⁻¹. ¹H NMR δ : 4.57 (1H, m, H-4), 3.69 (3H, m, H-5-6), 2.52 (2H, m, H-2), 2.23 (2H, m, H-3). ¹³C NMR δ : 178.0 (C-1), 80.7 (C-4), 73.5 (C-5), 63.3 (C-6), 28.4 (C-2), 23.9 (C-3).

(4R, 5R)-5-Hydroxy-6-para-toluenesulfonyloxy-4-hexanolide (14)

At -20°C to a solution of 13 (58 mg, 0.4 mmol) in CH₂Cl₂ (2mL) was added dropwise *para*-toluenesulfonyl chloride (75.3 mg, 0.4 mmol) in triethylamine (83 μ L) and dichloromethane (800 μ L). The mixture was stirred 3 hrs at -20°C, then overnight at room temperature and poured into water, extracted with CH₂Cl₂ and the combined organic layers were washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. Flash chromatography of the residue (CH₂Cl₂/AcOEt 8/2; Rf: 0.65 for the ditosyl and Rf: 0.21 for the monotosyl) afforded 10.6 mg of ditosyl product and 30.6 mg (26 %) of the monotosyl product 14.

¹H NMR δ : 8.8, 7.35 (4H, 2d, Ar, J=8Hz), 4.54 (1H, dt, H-4, J=2.7, 7Hz), 4.09 (2H, m, H-6), 3.91 (1H, m, H-5), 2.2-2.8 (5H, m, H-2, H-3, OH), 2.43 (3H, s, OCH₃).

(4R, 5R)-5-Hydroxy-6-triisopropylbenzensulfonyloxy-4-hexanolide (15)

At 0°C to a solution of 13 (58 mg, 0.4 mmol) in pyridine (900 μ L) was added dropwise triisopropyl benzenesulfonyl chloride (160.7 mg, 0.53 mmol). The mixture was stirred 45 min at 0°C and overnight at room temperature. Then CH₂Cl₂ and an aqueous hydrochloride acid solution were added. After extraction with CH₂Cl₂, the combined organic layers were washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. Flash chromatography of the residue (cyclohexane/AcOEt 1/1; Rf: 0.33) afforded 68 mg of 15 (41 %).

 1 H NMR δ : 7.2 (2H, s, Ar), 4.55 (1H, m, H-4), 3.8-4.3 (4H, m, H-5,-6, OH), 2.0-3.1 (7H, H-2, 3, CHMe₂), 1.3 (18H, m, CHMe₂).

(4R, 5R)-5, 6-Epoxy-4-hexanolide (16)

- a) From 13: Mitsunobu reaction on 13 was carried out under identical conditions as for 5 described beforehand. Flash chromatography (cyclohexane/AcOEt 2/8; Rf: 0.27) afforded 16 (65 % yield).
- b) From 15: At 0°C to a solution of 15 (250 mg, 0.6 mmol) in THF (6.5 mL) and dimethylsulfoxide (106 mL) was added NaH (17.4 mg, 0.73 mmol). After 2.5hrs stirring at room temperature, a saturated aqueous solution of NH₄OAc (2 mL) was added and the mixture was poured into a suspension of dichloromethane and brine. After extraction, the organic layers were dried (MgSO₄), filtered and concentrated

in vacuo to give an oil (65.7 mg) Flash chromatography of the oil (cyclohexane/AcOEt 2/8) afforded 49 mg of 1 6 (63 %).

¹H NMR δ : 4.53 (1H, m, H-4, J_{4,5}=3.4Hz, J_{3,4}= $J_{3',4}$ =6.6Hz), 3.08 (1H, m, H-5, J_{4,5}= $J_{5,6}$ = $J_{5,6}$ =3.4Hz), 2.71 (2H, m, H-6), 2.4-2.7 (2H, m, H-2), 2.2-2.4 (2H, m, H-3). ¹³C NMR δ : 176.5 (C-1), 77.2 (C-4), 53.1 (C-5), 43.8 (C-6), 27.7 (C-3), 24.9 (C-2).

(4R, 5R) (-)-Muricatacin (1)

- a) From **8**: To a solution of **8** (25.3 mg, 0.063 mmol) in CH_2Cl_2 (640 μL) was added at room temperature, water (37 μL) and dicyanodichloroquinone (21.5 mg, 0.095 mmol). After 2 hrs stirring, the mixture was poured into an aqueous saturated solution of NaHCO₃ and extracted with CH_2Cl_2 . Organic layers were washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. Flash chromatography of the residue afforded 13 mg of (-)- Muricatacin **1** (71 %).
- b) From 11: Decarbethoxylation of crude product 11 (obtained from 181 mg of 10) was carried out under identical conditions as for 7 described beforehand, and 41 mg of (-)-Muricatacin was obtained with 20 % yield from the epoxy alcohol 10 after flash chromatography (cyclohexane/AcOEt 7/3; Rf: 0.17).
- c) From 16: Nucleophilic opening of 16 (51 mg, 1.2 mmol) by undecyl magnesiumbromide in presence of Li₂CuCl₄ was carried out as for 3 described above but the mixture undecyl magnesiumbromide-Li₂CuCl₄ was poured at -78°C on the solution of the epoxylactone 16 in THF. After flash chromatography 34% of starting material 16 (20 mg) and 38% of (-)-Muricatacin 1 (49 mg) were obtained (58% yield of 1 based on 66% conversion of 16).
- 1 : m.p. 71°C, lit. 72°C, 9a 67-68°C, 9c 73°C, 9h 57-58°C, 8 [α]_D -22 (c 0.64, CHCl₃), -22 (c 0.67, MeOH), lit. -22.9 (c 1.1, CHCl₃), 9a -23.3 (CHCl₃), 8 -22.9 (c 1.1, CHCl₃), 9b -24.4 (c 1.70, MeOH), 9b -23.5(c 1, CHCl₃), 9d -18.8 (c 2.4, CHCl₃), 8 . IR (film) 3435, 1770 cm⁻¹.

¹H NMR (500 MHz) δ¹⁹: 4.39 (1H, part X highly coupled system ABCDXY, H-4, J_{4,5}=7.3Hz, J_{4,3β}=8Hz), 3.50 (1H, m, part Y, H-5, J_{4,5}=7.3Hz), 2.60 (1H, m, part AB, H-2α, J_{2α,3β}=9.9Hz, J_{3α,2α}=4.8Hz, J_{2α,2β}=-17.8Hz), 2.54 (1H, m, part AB, H-2β, J_{2β,3α}=9.2Hz, J_{2β,3β}=9.2Hz, J_{2α,2β}=-17.8Hz), 2.22 (1H, m, part CD, H-3α, J_{3α,4}=4.6Hz, J_{3α,2β}=9.2Hz, J_{3α,2α}=4.8Hz J_{3α,3β}=-12.6Hz), 2.11 (1H, m, part CD, H-3β, J_{3β,4}=8Hz, J_{3β,2β}=9.2Hz, J_{3β,2α}=9.9Hz, J_{3α,3β}=-12.6Hz), 1.92 (1H, d, OH), 1.51 (2H, m, H-6-6'), 1.23-1.37 (20H, m, (CH₂)₁₀), 0.86 (t, 3H, CH₃, J=6.7Hz).

 13 C NMR (125MHz) δ : 177.1 (C-1), 92.9 (C-4), 73.6 (C-5), 33.0 (C-6), 31.9 (C-7), 29.6, 29.5, 29.3, (C-8-15), 28.7, (C-2), 23.1 (C-3), 22.6 (C-16), 14.1 (C-17).

Anal. Calcd. for C₁₇H₃₂O₃: C, 71.79; H, 11.34. Found: C, 71.78; H, 11.37.

ACKNOWLEDGMENTS

We thank Dr. Josyane Gharbi-Benarous of our laboratory for the NMR discussions.

REFERENCES

- 1. Part II: see following paper in this issue.
- 2. Muller, C.J.; Kepner, R.T.; Webb, A.D. Am. J. Enol. Viticult. 1973, 24,5.
- 3. Muller, C.J.; Maggiora, L.; Kepner, R.E.; Webb, A.D. J. Agric. Food. Chem. 1969, 17, 1973.

- 4. Schumacher, J.N.; Green, C.R.; Best, F.W.; Newell, M.P. J. Agric. Food. Chem. 1977, 25, 310.
- 5. Wright, A.E.; Schäfer, M.; Miland, S.; Munnecke, D.E.; Sims, J.J. Tetrahedron Lett. 1989, 30, 5699.
- 6. Grafe, U.; Reinhardt, G.; Schade, W.; Krehes, D.; Eritt, I.; Fleck, W.F.; Heinrich, E.; Radics, L. J. Antibiotics 1982, 38, 609.
- 7. Rieser, M.J.; Kozlowski, U.F.; Wood, K.V.; McLaughin, J.L. Tetrahedron Lett. 1991, 32, 1137.
- 8. L. Factor: Kang, S.K.; Cho, H-S.; Sim, H-S.; Kim, B-K. J. Carbohydr. Chem. 1992, 11, 807 and references cited therein.
- Muricatacin: a) Scholz, G.; Tochtermann, W. Tetrahedron Lett. 1991, 32, 5535. b) Figadère, B.;
 Harmange, J-C.; Laurens, A.; Cavé, A. Tetrahedron Lett. 1991, 32, 7539. c) Marshall, J.A.;
 Welmaker, G.S. Synlett 1992, 537. d) Wang, Z.M.; Zhang, X.L.; Sharpless, K.B.; Sinha, S.C.;
 Sinha-Bazchi, A.; Keinan, E. Tetrahedron Lett.. 1992, 33, 6407. e) Tochtermann, W.; Scholz, G.;
 Bunte, G.; Wolf, C.; Peters, E.M.; Peters, K.; von Schnering, H.G. Liebigs Ann. Chem. 1992, 1069.
 f) Yao, Z.J.; Zhang, Y.B.; Wu, Y.L.; Huaxue Xuebao Acta Chimica Sinica 1992, 50, 901.
 g) Saïah, M.; Bessodes, M.; Antonakis, K. Tetrahedron Lett. 1993, 34, 1597. h) Makabe, H.; Tanaka, A.; Oritani, T. Biosei. Biotech. Biochem. 1993, 57, 807. i) Gravier-Pelletier, C.; Sanière, M.;
 Charvet, I.; Le Merrer, Y.; Depezay, J-C. Tetrahedron Lett. 1994, 35, 115.
- 10. Gravier-Pelletier, C.; Dumas, J.; Le Merrer, Y.; Depezay, J-C. J. Carbohydr. Chem. 1992, 11, 969.
- 11. Hicks, D.R.; Fraser-Reid, B. Synthesis 1974, 203.
- 12. a) Mitsunobu, O. Synthesis 1981, 1. b) Abushanab, E.; Vemishetti, P.; Leiby, R.W.; Singh, H.K.; Mikkilineni, A.B.; Wu, D.C.J.; Saibaba, R.; Panzica, R.P. J. Org. Chem. 1988, 53, 2598. c) For a recent review: Hughes, D.L. Organic Reactions 1992, 42, 335-640.
- 13. Takano, S.; Goto, E.; Hirama, M.; Ogasawara, K. Heterocycles 1981, 16, 381.
- 14. Horiba, K.; Yoshioka, T.; Tanaka, T.; Oikawa, Y.; Yonemitru, O. Tetrahedron 1986, 42, 3021.
- 15. Gravier-Pelletier, C.; Le Merrer, Y.; Depezay, J.C. Synthetic Commun. 1994, 24, 2843.
- 16. a) Dana, G.; Gharbi-Benarous, J.; Vaissermann, J.; Philoche-Levisalles, M.; Bois, C. *Tetrahedron* 1988, 44, 6357.
 - b) Gharbi-Benarous, J.; Dana, G.; Philoche-Levisalles, M.; Bois, C. Magn. Reson. Chem. 1988, 26, 457.
- a) Iwaki, S.; Marumo, S.; Saito, T.; Yamada, M.; Katagiri, K. J. Am. Chem. Soc. 1974, 96, 7842.
 b) Keinan, E.; Sinha, S.C.; Sinha-Bagchi, A.; Wang, Z.M.; Zhang, X.L.; Sharpless, K.B. Tetrahedron Lett. 1992, 33, 6411. c) Kang, S.K.; Kim, Y.S.; Lim, J.S.; Kim, S.G. Tetrahedron Lett. 1992, 32, 363 and references cited therein.
- 18. Wang, Z.H.; Qian, X.H.; Zhou, W.S. Tetrahedron Lett. 1990, 46, 1191.
- 19. The assignments have been made using 1D ¹H, ¹³C and 2D heteronuclear (F2 decoupled) NMR spectra on a Bruker AMX 500 spectrometer at room temperature (500 MHz for ¹H and 125 MHz for ¹³C). ¹H-¹H coupling constants were evaluated by simulation of the signals of H-2, H-3, H-4, H-5. The spectrum simulation was done on a Macintosh II computer using the software NMR II.